



Review

Autoimmune thyroid disorders



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ABSTRACT

Autoimmune thyroid diseases (AITD) result from a dysregulation of the immune system leading to an immune attack on the thyroid. AITD are T cell-mediated organ-specific autoimmune disorders. The prevalence of AITD is estimated to be 5%; however, the prevalence of antithyroid antibodies may be even higher. The AITD comprise two main clinical presentations: Graves' disease (GD) and Hashimoto's thyroiditis (HT), both characterized by lymphocytic infiltration of the thyroid parenchyma. The clinical hallmarks of GD and HT are thyrotoxicosis and hypothyroidism, respectively. The mechanisms that trigger the autoimmune attack to the thyroid are still under investigation. Epidemiological data suggest an interaction among genetic susceptibility and environmental triggers as the key factor leading to the breakdown of tolerance and the development of disease. Recent studies have shown the importance of cytokines and chemokines in the pathogenesis of AT and GD. In thyroid tissue, recruited T helper 1 (Th1) lymphocytes may be responsible for enhanced IFN- γ and TNF- α production, which in turn stimulates CXCL10 (the prototype of the IFN- γ -inducible Th1 chemokines) secretion from the thyroid cells, therefore creating an amplification feedback loop, initiating and perpetuating the autoimmune process. Associations exist between AITD and other organ specific (polyglandular autoimmune syndromes), or systemic autoimmune disorders (Sjögren's syndrome, rheumatoid arthritis, systemic lupus erythematosus, systemic sclerosis, cryoglobulinemia, sarcoidosis, psoriatic arthritis). Moreover, several studies have shown an association of AITD and papillary thyroid cancer. These data suggest that AITD patients should be accurately monitored for thyroid dysfunctions, the appearance of thyroid nodules, and other autoimmune disorders.

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1. Introduction

Autoimmune thyroid diseases (AITD) result from a dysregulation of the immune system leading to an immune attack on the thyroid. AITD are T cell-mediated organ-specific autoimmune disorders [1,2]. AITD are the most frequent autoimmune disorders, and the most common

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pathological conditions of the thyroid gland. The AITD comprise two main clinical presentations: Graves' disease (GD) and Hashimoto's thyroiditis (HT), that are both characterized by lymphocytic infiltration of the thyroid parenchyma. The clinical hallmarks of GD and HT are thyrotoxicosis and hypothyroidism, respectively.

The present study summarizes data about epidemiologic, risk factors and immunopathogenesis of HT.

1.1. Epidemiology

The prevalence of AITD is estimated to be 5% [3,4]; however, the prevalence of antithyroid antibodies (ATAs) without clinical disease may be even higher [5].

Studies [6–8] that have evaluated the changing epidemiology of HT have shown that: (1) women have a greater risk than men (about 4–10/1, female/male); (2) hypothyroidism from HT becomes more common with advancing age; (3) there are substantial geographic variability in the prevalence and incidence of HT; (4) prevalences of HT and thyroid antibodies differ with race; (5) ATA frequency increases with age, with a peak at around 45–55 years; and (6) populations that are iodine-sufficient have higher incidence of HT than those that are iodine-deficient.

In the Whickham study, the prevalence of spontaneous hypothyroidism from HT was 15/1000 in women, with mean age at diagnosis of 57 years, and less than 1/1000 in men [9]. The mean incidence of spontaneous hypothyroidism was 3.5/1000 in women and 0.6/1000 in men. Similar results have been recorded in other geographical areas [6].

The contemporary reported incidence rates of HT and hypothyroidism are higher than those in studies previously performed in similar regions [10].

However, it is not possible to know whether this is due to actual increased incidence or to the use of more accurate diagnostic procedures [11].

The HT is a prototypical organ-specific autoimmune disease. However, in many cases, AITD may be associated in the same patient with other organ-specific autoimmune attacks (such as in the case of type II autoimmune polyglandular syndrome), or less frequently with systemic autoimmune syndromes.

1.2. Risk factors

The mechanisms that trigger the autoimmune attack to the thyroid are still under investigation. Epidemiological data suggest an interaction among genetic susceptibility and environmental triggers as the key factor leading to the breakdown of tolerance and the development of disease.

1.3. Genetic susceptibility

Epidemiological evidence for a genetic susceptibility to AITD has been shown by the familial clustering of the disease (20–30% of AITD in siblings of affected patients), the sibling risk ratio (about 17) for AITD, and the increased prevalence of thyroid Abs (50%) of siblings of affected subjects [12]. The results of the twin studies show a concordance rate for AITD of 0.3–0.6 for monozygotic twins, compared to 0.00–0.1 for dizygotic twins [13,14]. From the twin studies, the heritability of GD has been calculated to be 79% in twin studies, and that of the presence of thyroid Abs was about 70% [14].

Several genes have been identified as significantly associated with the AITD and the presence of thyroid antibodies [15,16].

Among AITD genes detected by traditional case–control studies and tag single nucleotide polymorphism screening:

1. PTPN22 is involved in T-cell signal transduction through interaction with molecules essential for T-cell receptor signaling [17];
2. CTLA4 plays a role in inhibiting T-cell signaling [18];

3. Histocompatibility antigen (HLA) class II molecules play a key part in presenting exogenous antigens for recognition by CD4 + T-helper cells [19];
4. IL2RA encodes CD25 which is expressed on T-regulatory cells and is believed to be important in downregulating T-cell activity [20].

Other AITD genes were detected by case–control studies and confirmed by genome-wide association studies (GWAS):

1. FCRL3 is highly expressed during B-cell maturation and believed to both positively and negatively regulate B-cell signaling [21];
2. HLA class I molecules play a key role in presenting endogenous antigens, such a virally derived antigens, for recognition by CD8 + T cells [19];
3. TSHR is the receptor for TSH and is the primary autoantigenic target in GD [22].

Novel AITD genes have been detected by GWAS and ImmunoChip:

1. GDCG4p14 has been shown to be expressed in CD4 + T helper and CD8 + T cells [23];
2. BACH2 is expressed during B-cell maturation and is believed to control B-cell development and antibody production [24];
3. RNASET2 is expressed in CD4 + T-helper and CD8 + T cells [23];
4. FOXE1 is involved in thyroid gland morphogenesis and binds response elements in the thyroglobulin (Tg) and thyroid peroxidase promoters [25].

There are other AITD genes detected by GWAS and ImmunoChip whose function in AITD is currently unknown [15].

Interestingly among susceptibility genes whose function is known, 7/11 are involved in T cell function, strongly suggesting the importance of T cells in the immunopathogenesis of AITD.

1.4. Environmental factors

Environmental factors contribute to the occurrence of AITD for about 20%. Several environmental factors have been identified: radiation, iodine, smoking, infection, stress and drugs.

The link between environmental factors and autoimmunity is based on the principle that any injury resulting from infectious, chemical, radiological insults, may contribute to the activation of an innate immune response and, in susceptible individuals, to the development of AITD [26].

Radioiodine treatment of toxic goiter may be followed by the appearance of GD, even by Graves' ophthalmopathy (GO) [27]. Children exposed to radiation from Chernobyl showed a greater prevalence of thyroid autoantibody [28].

AITD tend to be more prevalent in areas with iodine sufficiency. Iodine supplementation of populations that were previously iodine deficient is associated with a transient increase of both autoimmune subclinical hypo- and hyperthyroidism [29].

Cigarette smoking has been associated with GD and with GO [30,31]. However, on the contrary, smoking decreases the risk of overt hypothyroidism as well as the prevalence of thyroid antibodies [32].

The thyroid is the organ with the highest selenium content because it expresses specific selenoproteins. After the discovery of myxoedematous cretinism following selenium repletion in iodine- and selenium-deficient children, many researches on links between thyroid and selenium have been published. Small amounts of selenium appear sufficient for adequate activity of deiodinases, however selenium status appears to have an impact on the development of thyroid pathologies. The importance of selenium supplementation in AITD has been emphasized [33].

Stress has been considered as a trigger factor for GD [34].

Among drugs, lithium treatment is associated with an increased prevalence of thyroid antibodies, hypothyroidism and, to a lesser extent, GD [35].

Thyroid autoimmunity might be relevant in amiodarone induced thyrotoxicosis [36].

The so-called “reconstitution Graves’ disease” has been observed in patients treated with anti-T cell anti-CD52 Campath monoclonal antibody for multilocal scleroderma, or in AIDS patients treated with antiretroviral therapy [37]. In both cases, GD occurs during the lymphocyte reconstitution phase suggesting the existence of an imbalance towards a Th2-mediated immune response.

Many studies have evaluated the contribution of viruses to the occurrence of AITD, mainly with no fully convincing or negative results [38].

1.5. HCV

However, recently, several studies have confirmed an association of hepatitis C virus (HCV) infection with AITD both in adults [39,40] and in children [41].

The thyroid disorders observed in patients with chronic hepatitis C (CHC) are characterized by a high risk of autoimmune thyroiditis (AT) and hypothyroidism in females, and high levels of anti-thyroperoxidase antibodies (AbTPO).

Recently, several studies have confirmed a high frequency of AT in patients with mixed cryoglobulinemia and hepatitis C (MC + HCV) and CHC.

In a case–control study serum AbTPO, and/or AbTg, and subclinical hypothyroidism were significantly more frequent in MC + HCV patients than in HCV-negative controls [42].

Moreover, a high prevalence of papillary thyroid cancer (PTC) has been observed in CHC patients, and more recently in MC + HCV patients, overall in the presence of AT [43–46].

The presence of HCV in the thyroid of chronically infected patients has been demonstrated [47,48].

More recently, it has been shown that HCV can infect a human thyroid cell line (ML1) in vitro. These findings suggest that HCV infection of thymocytes may play a role in the association between CHC and thyroid diseases [49].

Recent data have confirmed a strong association of AITD with interferon (IFN)- α therapy in patients with CHC. HCV and IFN- α can act in synergism to trigger AITD in patients. Approximately 40% of CHC patients develop thyroid disorders while receiving IFN- α . IFN-induced thyroiditis can manifest as destructive thyroiditis (or non-autoimmune hypothyroidism), or AT (with clinical features similar to those of GD or HT). IFN- α can induce thyroiditis via immune stimulation, such as direct toxic effects on the thyroid cells [50–52].

1.6. Endogenous factors

The marked predominance of AITD in female [53] suggests that estrogens have a significant role in AITD. The complex immunological changes associated with pregnancy and their postpartum regression are important factors. However, the female predisposition to AITD is also present in nulliparous women. Microchimerism, the presence of small populations of cells from one subject in another genetically distinct individual (microchimerism), has also been considered as one of the endogenous factors linked to AITD [14].

1.7. Immunopathogenesis

The common pathological feature of AITD is the presence of lymphocyte infiltrates within the thyroid. AT is characterized by lymphocytic infiltration, mainly of T cells, that may progressively replace thyroid tissue. Lymphoid infiltrates are also present in GD glands. Variable thyroid

follicular cell atrophy and fibrosis are characteristic of thyroiditis, while in GD thyroid follicles are hypertrophied [54].

CD8+ T cells are decreased in peripheral blood of patients with GD, HT and postpartum thyroiditis, just as in patients with other autoimmune diseases. Consequently, the CD4/CD8 ratio is increased. Also, activated T cells expressing HLA-DR are increased. In the thyroid tissue, T cell infiltrates associate CD4+ and CD8+ cells, often in the activated state. CD4+ may be predominant in Hashimoto glands [55].

B cell numbers are normal in circulation in AITD. In HT, B cells are found within the thyroid tissue, typically organized in secondary lymphoid follicles, sometimes with germinal centers. Intrathyroid B cells have been shown to produce antibody, suggesting that the thyroid is the main source of autoantibodies in vivo. Bone marrow and juxta-thyroid lymph node B cells are also a source of antibodies.

The prevalence of AbTg in patients with HT is 25–50%, while it is 90% for AbTPO [56]. In young patients with thyroiditis, however, AbTg may be present in the absence of AbTPO.

AbTg, predominantly IgG1 and 4, do not activate complement and are not pathogenic [57].

AbTPO are a sensitive marker of AITD, both in thyroiditis and GD. AbTPO are markers of thyroid dysfunction [58], in fact their presence is predictive of the subsequent occurrence of thyroid failure in AITD patients with subclinical hypothyroidism.

TSH receptor antibodies (TRAbs) are pathognomonic of GD. Indeed, the stimulating TRAbs are responsible for the hyperthyroidism of GD. TRAbs are present in more than 90% of the GD patients. TRAb radioimmunometric competition assays detect TRAbs through their capacity to bind to the TSH receptor; they provide no indication on the biological activity of the antibodies, stimulating, blocking, or neutral. The identification of the bioactivity of TRAbs requires a bioassay using cellular systems carrying functional TSH receptors [59]. Blocking antibodies are detected using a modified bioassay [60,61]. Blocking TRAbs are mostly detected in a fraction of patients with AT.

1.8. Cytokines, chemokines, and AITD

Recent studies have shown the importance of cytokines and chemokines in the pathogenesis of AT and GD. In thyroid tissue, recruited T helper 1 (Th1) lymphocytes may be responsible for enhanced IFN- γ and TNF- α production, which in turn stimulates CXCL10 (the prototype of the IFN- γ -inducible Th1 chemokines) secretion from the

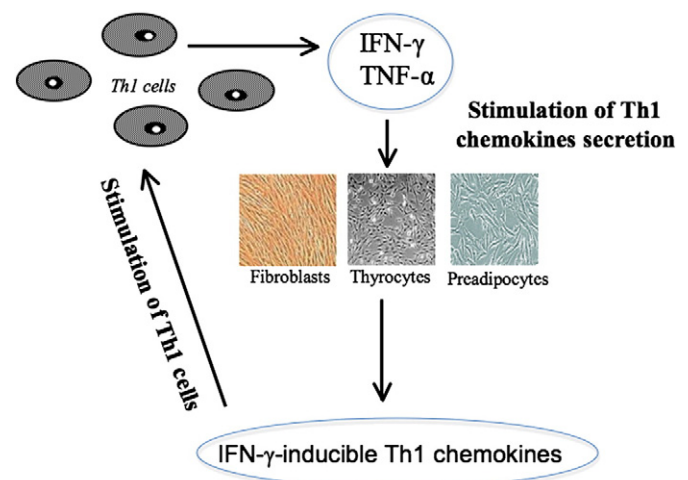


Fig. 1. In thyroid (and orbital tissue of GO patients), recruited Th1 lymphocytes may be responsible for enhanced IFN- γ and TNF- α production, which in turn stimulates CXCL10 (the prototype of the IFN- γ -inducible Th1 chemokines) secretion from the cells, therefore creating an amplification feedback loop, that initiates and perpetuates the autoimmune process.

thyroid cells, therefore creating an amplification feedback loop, initiating and perpetuating the autoimmune process (Fig. 1).

The IFN- γ -inducible protein 10 (IP-10/CXCL10) was initially identified as a chemokine that is induced by IFN- γ . CXCL10 exerts its function through binding to chemokine (C-X-C motif) receptor 3 (CXCR3) [62].

CXCL10 and its receptor, CXCR3, appear to contribute to the pathogenesis of many autoimmune diseases, organ specific [such as type 1 diabetes (T1D), GD and GO], or systemic [such as systemic lupus erythematosus (SLE), MC, Sjögren's syndrome (SS), sarcoidosis, psoriasis or systemic sclerosis (SSc)] [63–68].

The secretion of CXCL10 by (CD)4+, CD8+, and natural killer is dependent on IFN- γ . Under the influence of IFN- γ , and with synergism with TNF- α , CXCL10 is secreted by thyrocytes [69].

Determination of a high level of CXCL10 in peripheral fluids is therefore a marker of a Th1 orientated immune response [62].

High levels of circulating CXCL10 have been shown in patients with AT, in particular in the presence of a hypoechoic ultrasonographic pattern, which is a sign of a more severe lympho-monocytic infiltration, and in those with hypothyroidism [69].

For these reasons, it has been postulated that CXCL10 could be a marker of a stronger and more aggressive inflammatory response in the thyroid, subsequently leading to thyroid destruction and hypothyroidism. Further studies are needed to investigate whether CXCL10 is a novel therapeutic target in AT.

Moreover, CXCL10 appears to contribute to the pathogenesis of GD and GO (Fig. 1). Under the influence of IFN- γ , CXCL10 is secreted by thyrocytes (in GD), fibroblasts and preadipocytes (in GO) [70–72].

Circulating CXCL10 is associated with the active phase of GD in both newly diagnosed and relapsing hyperthyroid patients. Methimazole reduces CXCL10 secretion by isolated thyrocytes, decreases serum CXCL10 levels, and promotes a transition from Th1 to Th2 dominance in patients with GD active phase [73].

In GD patients the decrease of CXCL10 after thyroidectomy and radioiodine strongly suggests that this chemokine is mainly produced by the thyroid itself. In GO patients the increased concentrations of CXCL10, at least in part, reflect the activity of orbital inflammation [74,75].

A significant reduction in CXCL10 serum concentrations during corticosteroids and/or radiotherapy treatments, as compared both to control group and to basal values in GO patients, suggests that this chemokine could serve as a guideline in therapeutic decision-making in patients with GO [76,77].

Further studies are needed to evaluate whether CXCL10 is a novel therapeutic target in HT, GD and GO [78,79].

1.9. Other autoimmune diseases associated with thyroid autoimmunity

Associations exist between AITD and other organ specific, or systemic autoimmune disorders.

Among these syndromes, polyglandular autoimmune syndromes (PAS) are rare polyendocrinopathies characterized by the failure of several endocrine glands as well as nonendocrine organs, caused by an immune-mediated destruction of endocrine tissues.

In a study [80] of more than 15,000 adult patients with endocrine diseases, who have been screened, 360 patients with PAS have been found. T1D, GD, HT, Addison's disease, vitiligo, alopecia, hypogonadism, and pernicious anemia were observed in 61%, 33%, 33%, 19%, 20%, 6%, 5%, and 5%, respectively. The most common disease combination was T1D and AITD. In most patients, T1D was the first manifestation of PAS (48%).

Patients with PAS had significantly higher frequencies of the human leukocyte antigens A24, A31, B8, B51, B62, DR3, and DR4 (relative risk, 2.35, 2.74, 2.47, 7.17, 2.22, 1.94, and 2.46) vs. controls, and for A31, B15, B52, B55, DR2, DR11, and DR13 (relative risk, 2.51, 7.96, 3.99, 5.36, 4.46, 2.89, and 3.26) vs. T1D patients without PAS. This study suggests that patients with autoimmune endocrine disease should be

followed on a regular basis, if clinical disease is present, and serological measurement of organ-specific antibodies should follow [80].

The abovementioned study underlines the importance of a common genetic susceptibility in patients with AITD and T1D, as confirmed also in other studies [81].

Thyroid function abnormalities and thyroid autoantibodies have been frequently described in patients with systemic rheumatologic autoimmune diseases, such as SS, rheumatoid arthritis (RA), SLE and SSc [67,82,83].

A recent study showed that, despite contradictory results in the literature, there is a greater prevalence of the association between AITD and rheumatic diseases, highlighting the possibility of common pathogenic mechanisms among them [84].

Furthermore, more recently, a first study [85] has evaluated longitudinally the incidence of new cases of thyroid autoimmunity and dysfunction in 179 female patients with SSc, and 179 matched control subjects, with similar iodine intake (median follow-up of 73 months in patients with SSc vs. 94 months in control subjects). A high incidence of new cases of hypothyroidism, thyroid dysfunction, AbTPO positivity, and appearance of a hypoechoic thyroid pattern in sclerodermic patients (15.5, 21, 11, and 14.6 of 1000 patients per year; respectively) vs. those in control subjects were shown. A logistic regression analysis showed that in patients with SSc, the appearance of hypothyroidism was related to a borderline high initial TSH level, AbTPO positivity, and a hypoechoic and small thyroid. This study shows a high incidence of new cases of hypothyroidism and thyroid dysfunction in female sclerodermic patients, suggesting that these patients should have periodic thyroid function follow-up [85].

Furthermore, new associations of AITD are being uncovered for examples with MC, sarcoidosis, or psoriatic arthritis [86–88].

Many studies underline the importance of a common genetic susceptibility in patients with AITD and systemic autoimmunity.

The participation of the HLA of the haplotypes HLA-B8 and DR3 in both AITD and primary SS (pSS) has been suggested, because of the high frequency of those haplotypes in Caucasian patients with those diseases [89].

Genetic influence has been suggested in a study of 35 families with several cases of SLE concomitant with AITD, in which a gene of susceptibility was identified in 5q14.3–q15 (major locus of susceptibility for SLE, also found in AITD). That locus can be shared by patients with SLE and AITD, evidencing a potential genetic link between both diseases [90].

Another study has assessed the frequency of ATA and the genetic association with HLA class II antigens in 85 patients with scleroderma. Individuals with anti-TPO had a higher frequency of the HLA-DR15 allele than patients without those antibodies, suggesting that the HLA-DR15 allele can be a marker of immunogenicity for the formation of anti-TPO [91].

Even environmental factors could be implicated in the association of autoimmune disorders.

Recent reports that have shown that the serum and/or the tissue expressions of CXCL10 are increased in organ specific autoimmune diseases, such as AT, GD, T1D, and/or systemic rheumatological disorders like RA, SLE, SSc, MC, underline the importance of a common immunopathogenesis of these disorders, that are characterized by a Th1 prevalent autoimmune response in the initial, and/or active phases of these diseases [63,92,93].

1.10. The association of AITD and thyroid cancer

Several studies have shown an association of AITD and PTC [94].

A recent study [95] analyzed the frequency of PTC, TSH levels and thyroid autoantibodies in 13,738 patients [9824 untreated and 3914 under L-thyroxine (L-T(4))]. The frequency of PTC was significantly higher in nodular-HT than that in nodular goiter and was associated with increased levels of serum TSH. Treatment with L-T(4) reduces TSH levels and decreases the occurrence of clinically detectable PTC [95].

However, other studies have found that both thyroid autoimmunity and increased TSH represent independent risk factors for malignancy [96].

A high prevalence of PTC has been observed in CHC patients, and more recently in MC patients, overall in CHC or MC patients with AT [43,44,46].

The increased prevalence of PTC in AITD patients is clinically relevant since about 10–30% of these patients may have an aggressive disease, requiring systemic treatments [97–99].

2. Conclusion

Autoimmune thyroid diseases (AITD) result from a dysregulation of the immune system leading to an immune attack on the thyroid. AITD are T cell-mediated organ-specific autoimmune disorders. AITD are the most frequent autoimmune disorders. The AITD comprise two main clinical presentations: GD and HT, that are both characterized by lymphocytic infiltration of the thyroid parenchyma. The clinical hallmarks of GD and HT are thyrotoxicosis and hypothyroidism, respectively. The mechanisms that trigger the autoimmune attack to the thyroid are still under investigation. Epidemiological data suggest an interaction among genetic susceptibility and environmental triggers as the key factor leading to the breakdown of tolerance and the development of disease. Several environmental risk factors have been identified: radiation, iodine, selenium, smoking, infections, stress and drugs.

The common pathological feature of AITD is the presence of lymphocyte infiltrates within the thyroid. Recent studies have shown the importance of cytokines and chemokines in the pathogenesis of AT and GD. In thyroid tissue, recruited Th1 lymphocytes may be responsible for enhanced IFN- γ and TNF- α production, which in turn stimulates CXCL10 (the prototype of the IFN- γ -inducible Th1 chemokines) secretion from the thyroid cells, therefore creating an amplification feedback loop, initiating and perpetuating the autoimmune process.

Associations exist between AITD and other organ specific, or systemic autoimmune disorders [100]. Many studies underline the importance of a common genetic susceptibility in patients with AITD and systemic autoimmunity. Moreover, recent reports that have shown that the serum and/or the tissue expressions of CXCL10 are increased in organ specific autoimmune diseases, such as AT, GD, T1D, and/or systemic rheumatological disorders like RA, SLE, SSC, MC, underline the importance of a common immunopathogenesis of these disorders, that are characterized by a Th1 prevalent autoimmune response in the initial, and/or active phases of these diseases.

Several studies have found that both thyroid autoimmunity and increased TSH represent independent risk factors for thyroid malignancy.

The abovementioned data suggest that AITD patients should be accurately monitored, for thyroid dysfunctions, the appearance of thyroid nodules, and other organ specific or systemic autoimmune disorders during the course of the disease.

Take-home messages

- Autoimmune thyroid diseases (AITD) result from a dysregulation of the immune system leading to an immune attack on the thyroid and comprise two main clinical presentations: Graves' disease (GD) and Hashimoto's thyroiditis (HT). Epidemiological data suggest an interaction among genetic susceptibility and environmental triggers as the key factor leading to the breakdown of tolerance and the development of disease, and the importance of cytokines and chemokines in the pathogenesis of AT and GD has been shown.
- Associations exist between AITD and other organ specific or systemic autoimmune disorders, and papillary thyroid cancer. These data suggest that AITD patients should be accurately monitored, for thyroid dysfunctions, the appearance of thyroid nodules, and other autoimmune disorders.

Conflict of interest

The authors declare that they have no conflict of interest.

References

- [1] Romagnani S. The Th1/Th2 paradigm and allergic disorders. *Allergy* 1998;53:12–5.
- [2] Orgiazzi J. Thyroid autoimmunity. *Presse Med* 2012;41:e611–25.
- [3] Jacobson DL, Gange SJ, Rose NR, Graham NM. Epidemiology and estimated population burden of selected autoimmune diseases in the United States. *Clin Immunol Immunopathol* 1997;84:223–43.
- [4] Vanderpump MP, Tunbridge WM, French JM, Appleton D, Bates D, Clark F, et al. The incidence of thyroid disorders in the community: a twenty-year follow-up of the Whickham survey. *Clin Endocrinol (Oxf)* 1995;43:55–68.
- [5] Pearce SH, Leech NJ. Toward precise forecasting of autoimmune endocrinopathy. *J Clin Endocrinol Metab* 2004;89:544–7.
- [6] McLeod DS, Cooper DS. The incidence and prevalence of thyroid autoimmunity. *Endocrine* 2012;42:252–65.
- [7] McGrogan A, Seaman HE, Wright JW, de Vries CS. The incidence of autoimmune thyroid disease: a systematic review of the literature. *Clin Endocrinol (Oxf)* 2008; 69:687–96.
- [8] Vanderpump MP. The epidemiology of thyroid disease. *Br Med Bull* 2011;99: 39–51.
- [9] Tunbridge WM, Evered DC, Hall R, Appleton D, Brewis M, Clark F, et al. The spectrum of thyroid disease in a community: the Whickham survey. *Clin Endocrinol (Oxf)* 1977;7:481–9.
- [10] Pedersen IB, Laurberg P, Knudsen N, Jørgensen T, Perrild H, Ovesen L, et al. An increased incidence of overt hypothyroidism after iodine fortification of salt in Denmark: a prospective population study. *J Clin Endocrinol Metab* 2007;92: 3122–7.
- [11] Rizzo M, Rossi RT, Bonaffini O, Scisca C, Altavilla G, Calbo L, et al. Increased annual frequency of Hashimoto's thyroiditis between years 1988 and 2007 at a cytological unit of Sicily. *Ann Endocrinol (Paris)* 2010;71:525–34.
- [12] Tomer Y, Davies TF. Searching for the autoimmune thyroid disease susceptibility genes: from gene mapping to gene function. *Endocr Rev* 2003;24:694–717.
- [13] Hansen PS1, Brix TH, Iachine I, Kyvik KO, Hegedus L. The relative importance of genetic and environmental effects for the early stages of thyroid autoimmunity: a study of healthy Danish twins. *Eur J Endocrinol* 2006;154:29–38.
- [14] Brix TH, Hegedus L. Twin studies as a model for exploring the aetiology of autoimmune thyroid disease. *Clin Endocrinol* 2012;76:457–64.
- [15] Simmonds MJ. GWAS in autoimmune thyroid disease: redefining our understanding of pathogenesis. *Nat Rev Endocrinol* 2013;9:277–87.
- [16] Tomer Y, Ban Y, Concepcion E, Barbesino G, Villanueva R, Greenberg DA, et al. Common and unique susceptibility loci in Graves and Hashimoto diseases: results of whole-genome screening in a data set of 102 multiplex families. *Am J Hum Genet* 2003;73:736–47.
- [17] Smyth D, Cooper JD, Collins JE, Heward JM, Franklyn JA, Howson JM, et al. Replication of an association between the lymphoid tyrosine phosphatase locus (LYP/PTPN22) with type 1 diabetes, and evidence for its role as a general autoimmunity locus. *Diabetes* 2004;53:3020–3.
- [18] Ueda H, Howson JM, Esposito L, Heward J, Snook H, Chamberlain G, et al. Association of the T-cell regulatory gene CTLA4 with susceptibility to autoimmune disease. *Nature* 2003;423:506–11.
- [19] Gough SC, Simmonds MJ. The HLA region and autoimmune disease: associations and mechanisms of action. *Curr Genomics* 2007;8:453–65.
- [20] Lowe CE, Cooper JD, Brusko T, Walker NM, Smyth DJ, Bailey R, et al. Large-scale genetic fine mapping and genotype-phenotype associations implicate polymorphism in the IL2RA region in type 1 diabetes. *Nat Genet* 2007;39:1074–82.
- [21] Kochi Y, Yamada R, Suzuki A, Harley JB, Shirasawa S, Sawada T, et al. A functional variant in FCRL3, encoding Fc receptor-like 3, is associated with rheumatoid arthritis and several autoimmunities. *Nat Genet* 2005;37:478–85.
- [22] Brand OJ, Barrett JC, Simmonds MJ, Newby PR, McCabe CJ, Bruce CK, et al. Association of the thyroid stimulating hormone receptor gene (TSHR) with Graves' disease. *Hum Mol Genet* 2009;18:1704–13.
- [23] Chu X, Pan CM, Zhao SX, Liang J, Gao GQ, Zhang XM, et al. China Consortium for Genetics of Autoimmune Thyroid Disease. A genome-wide association study identifies two new risk loci for Graves' disease. *Nat Genet* 2011;43:897–901.
- [24] Simmonds MJ. Evaluating the role of B Cells in autoimmune disease: more than just initiators of disease? In: Berhardt LV, editor. *Advances in medicine and biology* New York: Nova Science Publisher, Inc.; 2011. p. 151–76.
- [25] Castanet M, Polak M. Spectrum of human Foxe1/TF2 mutations. *Horm Res Paediatr* 2010;73:423–9.
- [26] Kawashima A, Tanigawa K, Akama T, Yoshihara A, Ishii N, Suzuki K. Innate immune activation and thyroid autoimmunity. *J Clin Endocrinol Metab* 2011;96:3661–71.
- [27] Dunkelmann S, Wolf R, Koch A, Kittner C, Groth P, Schuemichen C. Incidence of radiation-induced Graves' disease in patients treated with radioiodine for thyroid autonomy before and after introduction of a high-sensitivity TSH receptor antibody assay. *Eur J Nucl Med Mol Imaging* 2004;31:1428–34.
- [28] Agate L, Mariotti S, Elisei R, Mossa P, Pacini F, Molinaro E, et al. Thyroid autoantibodies and thyroid function in subjects exposed to Chernobyl fallout during childhood: evidence for a transient radiation-induced elevation of serum thyroid antibodies without an increase in thyroid autoimmune disease. *J Clin Endocrinol Metab* 2008;93:2729–36.

- [29] Laurberg P, Jørgensen T, Perrild H, Ovesen L, Knudsen N, Pedersen IB, et al. The Danish investigation on iodine intake and thyroid disease, DanThyr: status and perspectives. *Eur J Endocrinol* 2006;155:219–28.
- [30] Prummel MF, Wiersinga WM. Smoking and risk of Graves' disease. *JAMA* 1993; 269:479–82.
- [31] Bartalena L, Marcocci C, Tanda ML, Manetti L, Dell'Unto E, Bartolomei MP, et al. Cigarette smoking and treatment outcomes in Graves ophthalmopathy. *Ann Intern Med* 1998;129:632–5.
- [32] Carlé A, Bülow Pedersen I, Knudsen N, Perrild H, Ovesen L, Banke Rasmussen L, et al. Smoking cessation is followed by a sharp but transient rise in the incidence of overt autoimmune hypothyroidism — a population-based, case–control study. *Clin Endocrinol (Oxf)* 2012;77:764–72.
- [33] Drutel A, Archambeaud F, Caron P. Selenium and the thyroid gland: more good news for clinicians. *Clin Endocrinol (Oxf)* 2013;78:155–64.
- [34] Mizokami T, Wu Li A, El-Kaissi S, Wall JR. Stress and thyroid autoimmunity. *Thyroid* 2004;14:1047–55.
- [35] Lazarus JH. Lithium and thyroid. *Best Pract Res Clin Endocrinol Metab* 2009;23: 723–33.
- [36] Martino E, Macchia E, Aghini-Lombardi F, Antonelli A, Lenziardi M, Concetti R, et al. Is humoral thyroid autoimmunity relevant in amiodarone iodine-induced thyrotoxicosis (AITT)? *Clin Endocrinol (Oxf)* 1986;24:627–33.
- [37] Weetman AP. Immune reconstitution syndrome and the thyroid. *Best Pract Res Clin Endocrinol Metab* 2009;23:693–702.
- [38] Desailoud R, Hober D. Viruses and thyroiditis: an update. *Virology* 2009;6:5.
- [39] Antonelli A, Ferri C, Fallahi P, Ferrari SM, Ghinoi A, Rotondi M, et al. Thyroid disorders in chronic hepatitis C virus infection. *Thyroid* 2006;16:563–72.
- [40] Giordano TP, Henderson L, Landgren O, Chiao EY, Kramer JR, El-Serag H, et al. Risk of non-Hodgkin lymphoma and lymphoproliferative precursor diseases in US veterans with hepatitis C virus. *JAMA* 2007;297:2010–7.
- [41] Indolfi G, Stagi S, Bartolini E, Salti R, de Martino M, Azzari C, et al. Thyroid function and anti-thyroid autoantibodies in untreated children with vertically acquired chronic hepatitis C virus infection. *Clin Endocrinol (Oxf)* 2008;68:117–21.
- [42] Antonelli A, Ferri C, Fallahi P, Giuggioli D, Nesti C, Longombardo G, et al. Thyroid involvement in patients with overt HCV-related mixed cryoglobulinemia. *QJM* 2004;97:499–506.
- [43] Antonelli A, Ferri C, Fallahi P, Pampana A, Ferrari SM, Barani L, et al. Thyroid cancer in HCV-related chronic hepatitis patients: a case–control study. *Thyroid* 2007;17: 447–51.
- [44] Montella M, Crispo A, de Bellis G, Izzo F, Frigeri F, Ronga D, et al. HCV and cancer: a case–control study in a high-endemic area. *Liver* 2001;21:335–41.
- [45] Ferri C, Antonelli A, Mascia MT, Sebastiani M, Fallahi P, Ferrari D, et al. B-cells and mixed cryoglobulinemia. *Autoimmun Rev* 2007;7:114–20.
- [46] Antonelli A, Ferri C, Fallahi P, Nesti C, Zignego AL, Maccheroni M. Thyroid cancer in HCV-related mixed cryoglobulinemia patients. *Clin Exp Rheumatol* 2002;20:693–6.
- [47] Gowans EJ. Distribution of markers of hepatitis C virus infection throughout the body. *Semin Liver Dis* 2000;20:85–102.
- [48] Bartolomé J, Rodríguez-Iñigo E, Quadros P, Vidal S, Pascual-Miguelañez I, Rodríguez-Montes JA, et al. Detection of hepatitis C virus in thyroid tissue from patients with chronic HCV infection. *J Med Virol* 2008;80:1588–94.
- [49] Blackard JT, Kong L, Huber AK, Tomer Y. Hepatitis C virus infection of a thyroid cell line: implications for pathogenesis of hepatitis C virus and thyroiditis. *Thyroid* 2013;23:863–70.
- [50] Menconi F, Hasham A, Tomer Y. Environmental triggers of thyroiditis: hepatitis C and interferon- α . *J Endocrinol Invest* 2011;34:78–84.
- [51] Ferri C, Antonelli A, Mascia MT, Sebastiani M, Fallahi P, Ferrari D, et al. HCV-related autoimmune and neoplastic disorders: the HCV syndrome. *Dig Liver Dis* 2007;39:13–21.
- [52] Villa E, Karampatou A, Cammà C, Di Leo A, Luongo M, Ferrari A, et al. Early menopause is associated with lack of response to antiviral therapy in women with chronic hepatitis C. *Gastroenterology* 2011;140:818–29.
- [53] Fish EN. The X-files in immunity: sex-based differences predispose immune responses. *Nat Rev Immunol* 2008;8:737–44.
- [54] Armengol MP, Juan M, Lucas-Martín A, Fernández-Figuera MT, Jaraquemada D, Gallard T, et al. Thyroid autoimmune disease: demonstration of thyroid antigen-specific B cells and recombination-activating gene expression in chemokine-containing active intrathyroidal germinal centers. *Am J Pathol* 2001;159:861–73.
- [55] McIntosh RS, Tandon N, Pickerill AP, Davies R, Barnett D, Weetman AP. IL-2 receptor-positive intrathyroidal lymphocytes in Graves' disease. Analysis of V alpha transcript microheterogeneity. *J Immunol* 1993;151:3884–93.
- [56] Pearce EN, Farwell AP, Braverman LE. Thyroiditis. *N Engl J Med* 2003;348:2646–55.
- [57] Huber G, Staub JJ, Meier C, Mittrache C, Guglielmetti M, Huber P, et al. Prospective study of the spontaneous course of subclinical hypothyroidism: prognostic value of thyrotropin, thyroid reserve, and thyroid antibodies. *J Clin Endocrinol Metab* 2002;87:3221–6.
- [58] McLachlan SM, Rapoport B. Thyroid peroxidase as an autoantigen. *Thyroid* 2007; 17:939–48.
- [59] Costagliola S, Morgenthaler NG, Hoermann R, Badenhoop K, Struck J, Freitag D, et al. Second generation assay for thyrotropin receptor antibodies has superior diagnostic sensitivity for Graves' disease. *J Clin Endocrinol Metab* 1999;84:90–7.
- [60] Madec AM, Clavel S, Stefanutti A, Orgiazzi J. Blocking anti-thyrotropin receptor antibodies desensitize cultured human thyroid cells. *Endocrinology* 1988;123:2062–6.
- [61] Li Y, Kim J, Diana T, Klases R, Olivo PD, Kahaly GJ. A novel bioassay for anti-thyrotropin receptor autoantibodies detects both thyroid-blocking and stimulating activity. *Clin Exp Immunol* 2013;173:390–7.
- [62] Antonelli A, Rotondi M, Fallahi P, Ferrari SM, Paolicchi A, Romagnani P, et al. Increase of CXC chemokine CXCL10 and CC chemokine CCL2 serum levels in normal ageing. *Cytokine* 2006;34:32–8.
- [63] Antonelli A, Ferrari SM, Giuggioli D, Ferrannini E, Ferri C, Fallahi P. Chemokine (C-X-C motif) ligand (CXCL)10 in autoimmune diseases. *Autoimmun Rev* 2014; 13:272–80.
- [64] Antonelli A, Baj G, Marchetti P, Fallahi P, Surico N, Pupilli C, et al. Human anti-CD38 autoantibodies raise intracellular calcium and stimulate insulin release in human pancreatic islets. *Diabetes* 2001;50:985–91.
- [65] Antonelli A, Fallahi P, Delle Sedie A, Ferrari SM, Maccheroni M, Bombardieri S, et al. High values of Th1 (CXCL10) and Th2 (CCL2) chemokines in patients with psoriatic arthritis. *Clin Exp Rheumatol* 2009;27:22–7.
- [66] Antonelli A, Ferri C, Fallahi P, Ferrari SM, Sebastiani M, Ferrari D, et al. High values of CXCL10 Serum levels in mixed cryoglobulinemia associated with hepatitis C infection. *Am J Gastroenterol* 2008;103:2488–94.
- [67] Antonelli A, Ferri C, Fallahi P, Cazzato M, Ferrari SM, Sebastiani M, et al. Clinical and subclinical autoimmune thyroid disorders in systemic sclerosis. *Eur J Endocrinol* 2007;156:431–7.
- [68] Antonelli A, Ferri C, Fallahi P, Colaci M, Giuggioli D, Ferrari SM, et al. Th1 and Th2 chemokine serum levels in systemic sclerosis in the presence or absence of autoimmune thyroiditis. *J Rheumatol* 2008;35:1809–11.
- [69] Ruffilli I, Ferrari SM, Colaci M, Ferri C, Fallahi P, Antonelli A. IP-10 in autoimmune thyroiditis. *Horm Metab Res* 2014. <http://dx.doi.org/10.1055/s-0034-1382053>.
- [70] Antonelli A, Rotondi M, Ferrari SM, Fallahi P, Romagnani P, Franceschini SS, et al. Interferon-gamma-inducible alpha-chemokine CXCL10 involvement in Graves' ophthalmopathy: modulation by peroxisome proliferator-activated receptor-gamma agonists. *J Clin Endocrinol Metab* 2006;91:614–20.
- [71] Antonelli A, Ferrari SM, Fallahi P, Frascerra S, Santini E, Franceschini SS, et al. Monokine induced by interferon gamma (IFN γ) (CXCL9) and IFN γ inducible T-cell alpha-chemoattractant (CXCL11) involvement in Graves' disease and ophthalmopathy: modulation by peroxisome proliferator-activated receptor-gamma agonists. *J Clin Endocrinol Metab* 2009;94:1803–9.
- [72] Antonelli A, Ferrari SM, Frascerra S, Pupilli C, Mancusi C, Metelli MR, et al. CXCL9 and CXCL11 chemokines modulation by peroxisome proliferator-activated receptor-alpha agonists secretion in Graves' and normal thyrocytes. *J Clin Endocrinol Metab* 2010;95:413–20.
- [73] Antonelli A, Rotondi M, Fallahi P, Romagnani P, Ferrari SM, Barani L, et al. Increase of interferon-gamma-inducible CXC chemokine CXCL10 serum levels in patients with active Graves' disease, and modulation by methimazole therapy. *Clin Endocrinol (Oxf)* 2006;64:189–95.
- [74] Antonelli A, Rotondi M, Fallahi P, Grosso M, Boni G, Ferrari SM, et al. Iodine-131 given for therapeutic purposes modulates differently interferon-gamma-inducible alpha-chemokine CXCL10 serum levels in patients with active Graves' disease or toxic nodular goiter. *J Clin Endocrinol Metab* 2007;92:1485–90.
- [75] Antonelli A, Fallahi P, Rotondi M, Ferrari SM, Serio M, Miccoli P. Serum levels of the interferon-gamma-inducible alpha chemokine CXCL10 in patients with active Graves' disease, and modulation by methimazole therapy and thyroidectomy. *Br J Surg* 2006;93:1226–31.
- [76] Zhu W, Ye L, Shen L, Jiao Q, Huang F, Han R, et al. A prospective, randomized trial of intravenous glucocorticoids therapy with different protocols for patients with Graves' ophthalmopathy. *J Clin Endocrinol Metab* 2014;99:1999–2007.
- [77] Mysliwiec J, Palyga I, Kosciuszko M, Kowalska A, Gorska M. Circulating CXCL9 and CXCL10 as markers of activity of Graves' orbitopathy during treatment with corticosteroids and teloradiotherapy. *Horm Metab Res* 2012;44:957–61.
- [78] Fallahi P, Ferrari SM, Corrado A, Giuggioli D, Ferri C, Antonelli A. Targeting chemokine (C-X-C motif) receptor 3 in thyroid autoimmunity. *Recent Patents Endocr Metab Immune Drug Discov* 2014;8:95–101.
- [79] Lee SH, Lim SY, Choi JH, Jung JC, Oh S, Kook KH, et al. Benzylideneacetophenone derivatives attenuate IFN- γ -induced IP-10/CXCL10 production in orbital fibroblasts of patients with thyroid-associated ophthalmopathy through STAT-1 inhibition. *Exp Mol Med* 2014;46:100.
- [80] Dittmar M, Kahaly GJ. Polyglandular autoimmune syndromes: immunogenetics and long-term follow-up. *J Clin Endocrinol Metab* 2003;88:2983–92.
- [81] Antonelli A, Tuomi T, Nannipieri M, Fallahi P, Nesti C, Okamoto H, et al. Autoimmunity to CD38 and GAD in Type I and Type II diabetes: CD38 and HLA genotypes and clinical phenotypes. *Diabetologia* 2002;45:1298–306.
- [82] Antonelli A, Fallahi P, Mosca M, Ferrari SM, Ruffilli I, Corti A, et al. Prevalence of thyroid dysfunctions in systemic lupus erythematosus. *Metabolism* 2010;59:896–900.
- [83] Antonelli A, Mosca M, Fallahi P, Neri R, Ferrari SM, D'Ascanio A, et al. Thyroid cancer in systemic lupus erythematosus: a case–control study. *J Clin Endocrinol Metab* 2010;95:314–8.
- [84] Robazzi TC, Adan LF. Autoimmune thyroid disease in patients with rheumatic diseases. *Rev Bras Reumatol* 2012;52:417–30.
- [85] Antonelli A, Fallahi P, Ferrari SM, Mancusi C, Giuggioli D, Colaci M, et al. Incidence of thyroid disorders in systemic sclerosis: results from a longitudinal follow-up. *J Clin Endocrinol Metab* 2013;98:1198–202.
- [86] Antonelli A, Fazzi P, Fallahi P, Ferrari SM, Ferrannini E. Prevalence of hypothyroidism and Graves disease in sarcoidosis. *Chest* 2006;130:526–32.
- [87] Antonelli A, Delle Sedie A, Fallahi P, Ferrari SM, Maccheroni M, Ferrannini E, et al. High prevalence of thyroid autoimmunity and hypothyroidism in patients with psoriatic arthritis. *J Rheumatol* 2006;33:2026–8.
- [88] Fallahi P, Ferrari SM, Giuggioli D, Manfredi A, Mancusi C, Fabiani S, et al. Thyroid involvement in hepatitis C-associated mixed cryoglobulinemia. *Hormones (Athens)* 2014;13:16–23.
- [89] Alfariis N, Curiel R, Tabbara S, Irwing MS. Autoimmune thyroid disease and Sjögren syndrome. *J Clin Rheumatol* 2010;16:146–7.
- [90] Namjou B, Kelly JA, Kilpatrick J, Kaufman KM, Nath SK, Scofield RH, et al. Linkage at 5q14.3–15 in multiplex systemic lupus erythematosus pedigrees stratified by autoimmune thyroid disease. *Arthritis Rheum* 2005;52:3646–50.

- [91] Molteni M, Barili M, Eisera N, Scrofani S, Mascagni B, Zulian C, et al. Anti-thyroid antibodies in Italian scleroderma patients: association of anti-thyroid peroxidase (anti-TPO) antibodies with HLA-DR15. *Clin Exp Rheumatol* 1997;15:529–34.
- [92] Lee EY, Lee ZH, Song YW. CXCL10 and autoimmune diseases. *Autoimmun Rev* 2009;8:379–83.
- [93] Antonelli A, Ferrari SM, Corrado A, Ferrannini E, Fallahi P. CXCR3, CXCL10 and type 1 diabetes. *Cytokine Growth Factor Rev* 2014;25:57–65.
- [94] Boi F, Lai ML, Marziani B, Minerba L, Faa G, Mariotti S. High prevalence of suspicious cytology in thyroid nodules associated with positive thyroid autoantibodies. *Eur J Endocrinol* 2005;153:637–42.
- [95] Fiore E, Rago T, Latrofa F, Provenzale MA, Piaggi P, Delitala A, et al. Hashimoto's thyroiditis is associated with papillary thyroid carcinoma: role of TSH and of treatment with L-thyroxine. *Endocr Relat Cancer* 2011;18:429–37.
- [96] Boi F, Minerba L, Lai ML, Marziani B, Figus B, Spanu F, et al. Both thyroid autoimmunity and increased serum TSH are independent risk factors for malignancy in patients with thyroid nodules. *J Endocrinol Invest* 2013;36:313–20.
- [97] Neri S, Boraschi P, Antonelli A, Falaschi F, Baschieri L. Pulmonary function, smoking habits, and high resolution computed tomography (HRCT) early abnormalities of lung and pleural fibrosis in shipyard workers exposed to asbestos. *Am J Ind Med* 1996;30:588–95.
- [98] Antonelli A, Bocci G, La Motta C, Ferrari SM, Fallahi P, Fioravanti A, et al. Novel pyrazolopyrimidine derivatives as tyrosine kinase inhibitors with antitumoral activity in vitro and in vivo in papillary dedifferentiated thyroid cancer. *J Clin Endocrinol Metab* 2011;96:288–96.
- [99] Antonelli A, Fallahi P, Ferrari SM, Carpi A, Berti P, Materazzi G, et al. Dedifferentiated thyroid cancer: a therapeutic challenge. *Biomed Pharmacother* 2008;62:559–63.
- [100] Weetman AP. Diseases associated with thyroid autoimmunity: explanations for the expanding spectrum. *Clin Endocrinol (Oxf)* 2011;74:411–8.

Neutrophil-mediated IFN activation in the bone marrow alters B cell development in human and murine systemic lupus erythematosus.

In appropriate activation of type I IFN plays a key role in the pathogenesis of autoimmune disease, including systemic lupus erythematosus (SLE). In this study, **Palanichamy A. et al (J Immunol. 2014; 192(3): 906-18)** report on the presence of IFN activation in SLE bone marrow (BM), as measured by an IFN gene signature, increased IFN regulated chemokines, and direct production of IFN by BM-resident cells, associated with profound changes in B cell development. The majority of SLE patients had an IFN signature in the BM that was more pronounced than the paired peripheral blood and correlated with both higher autoantibodies and disease activity. Pronounced alterations in B cell development were noted in SLE in the presence of an IFN signature with a reduction in the fraction of pro/pre-B cells, suggesting an inhibition in early B cell development and an expansion of B cells at the transitional stage. These B cell changes strongly correlated with an increase in BAFF and APRIL expression in the IFN-high BM. Furthermore, they found that BM neutrophils in SLE were prime producers of IFN- α and B cell factors. In NZM lupus-prone mice, similar changes in B cell development were observed and mediated by IFN, given abrogation in NZM mice lacking type-I IFNR. BM neutrophils were abundant, responsive to, and producers of IFN, in close proximity to B cells. These results indicate that the BM is an important but previously unrecognized target organ in SLE with neutrophil-mediated IFN activation and alterations in B cell ontogeny and selection.